



SCHOOL OF PHARMACY

UNIVERSITY of WASHINGTON

*Pursuing the holy grail of predicting and verifying
tissue drug concentrations: A proteomics and PET
imaging approach*

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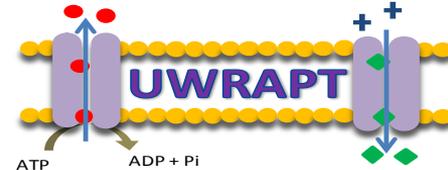
University of Washington

Seattle, WA

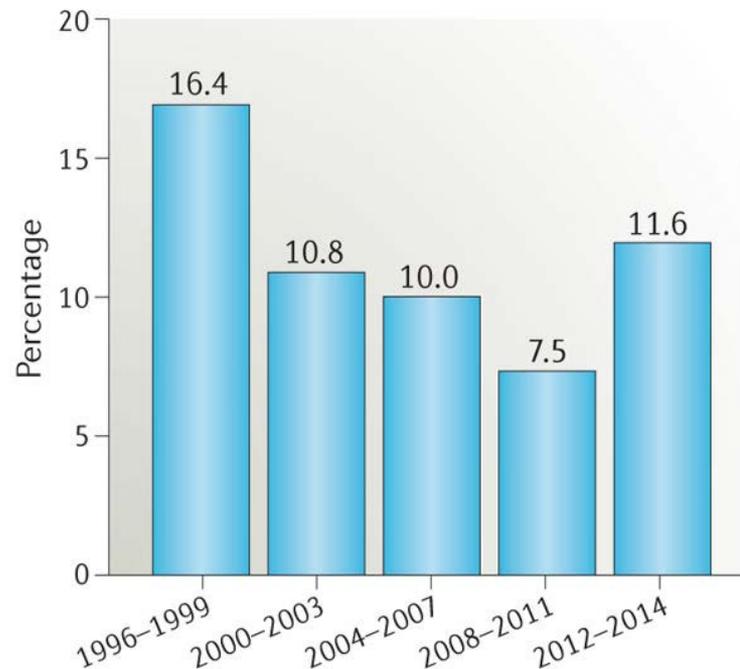
jash@uw.edu



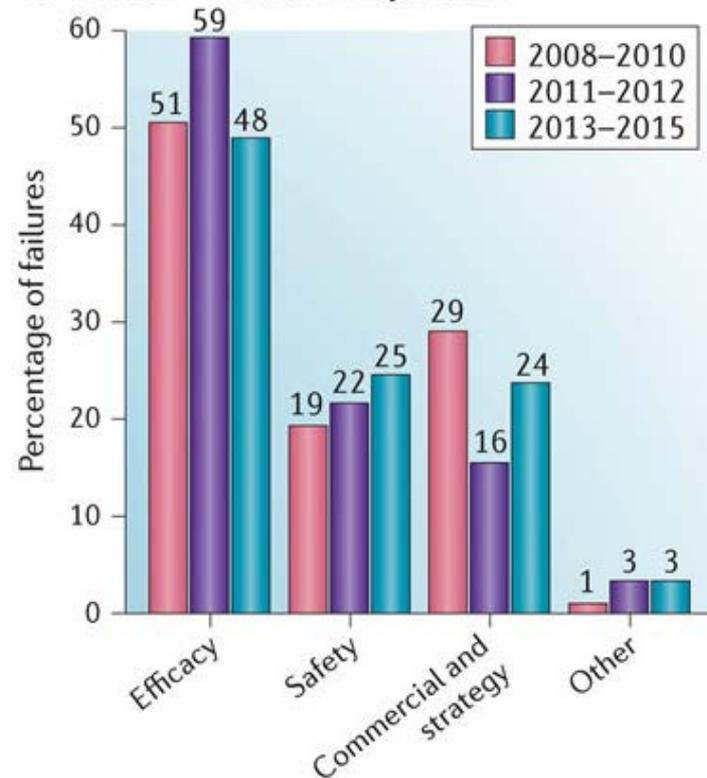
Failure Rate and Reasons for Failure in Drug Development



b Cumulative success rate Phase I to launch
Percentage likelihood of moving from Phase I to launch



b Reason for failure in phase II

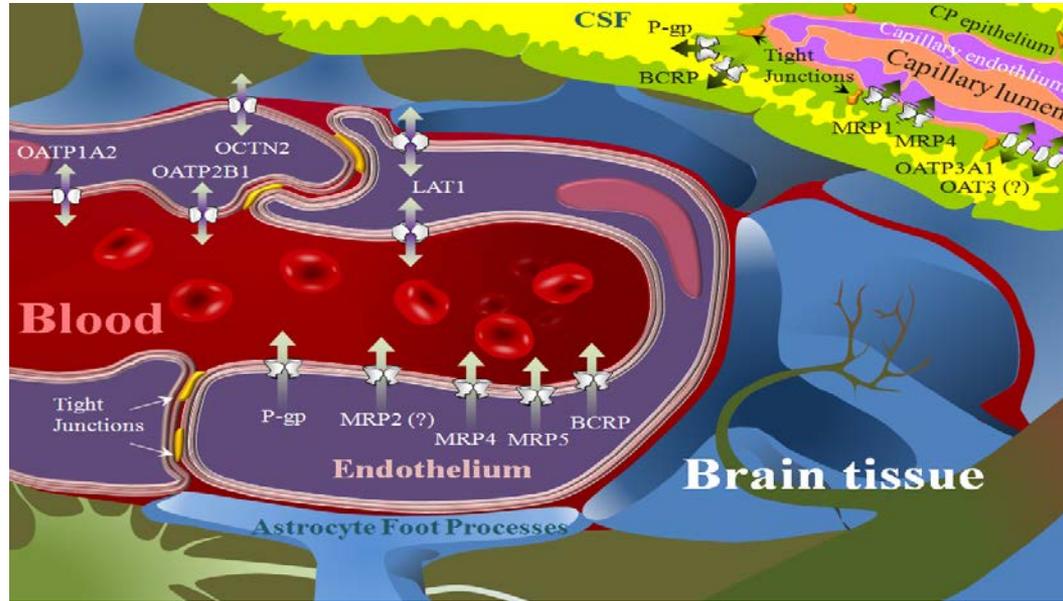
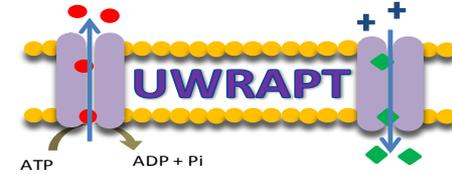


One Possible Reason for Lack of Drug Efficacy & Safety



- Unable to measure or predict tissue conc. of drugs
- Unbound plasma conc. \neq unbound tissue conc. if transporters are involved, i.e. asymmetry between blood and tissue drug conc. (e.g. liver:blood due to OATPs)

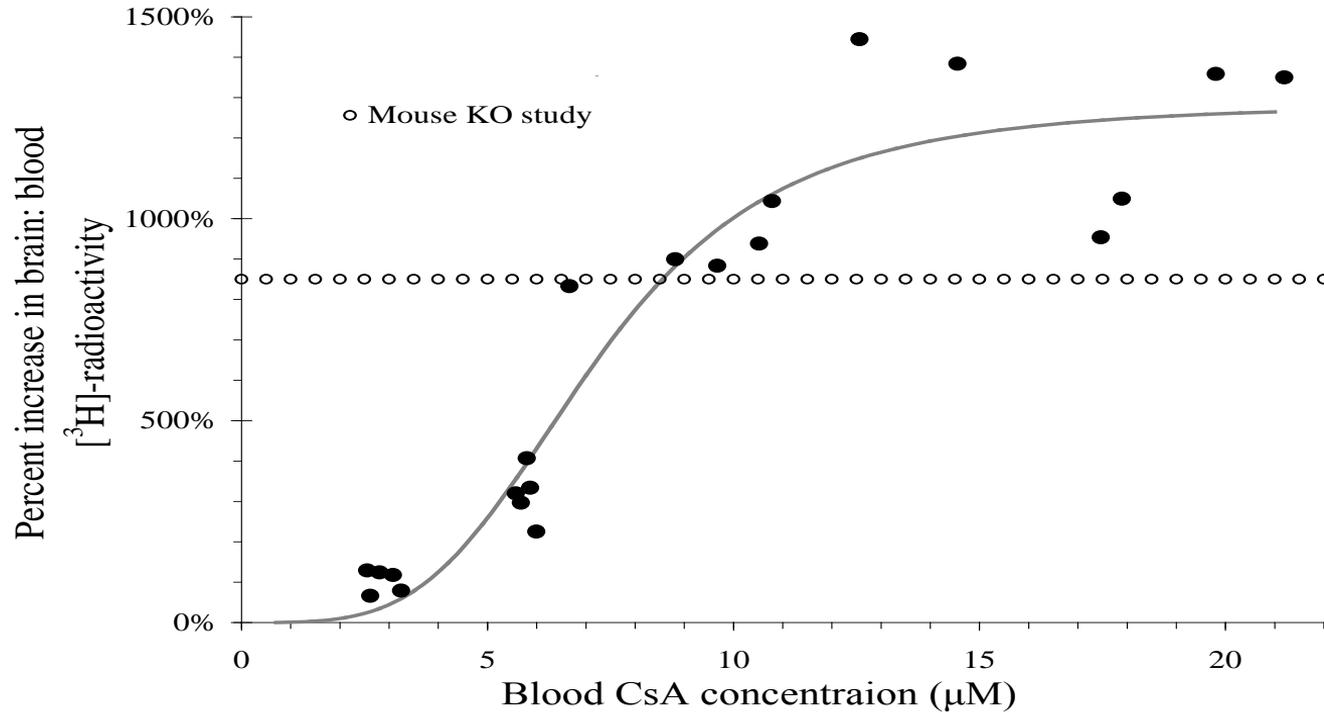
Asymmetry In Brain: Blood Drug Conc. Due to Blood-Brain Barrier Efflux Transporters



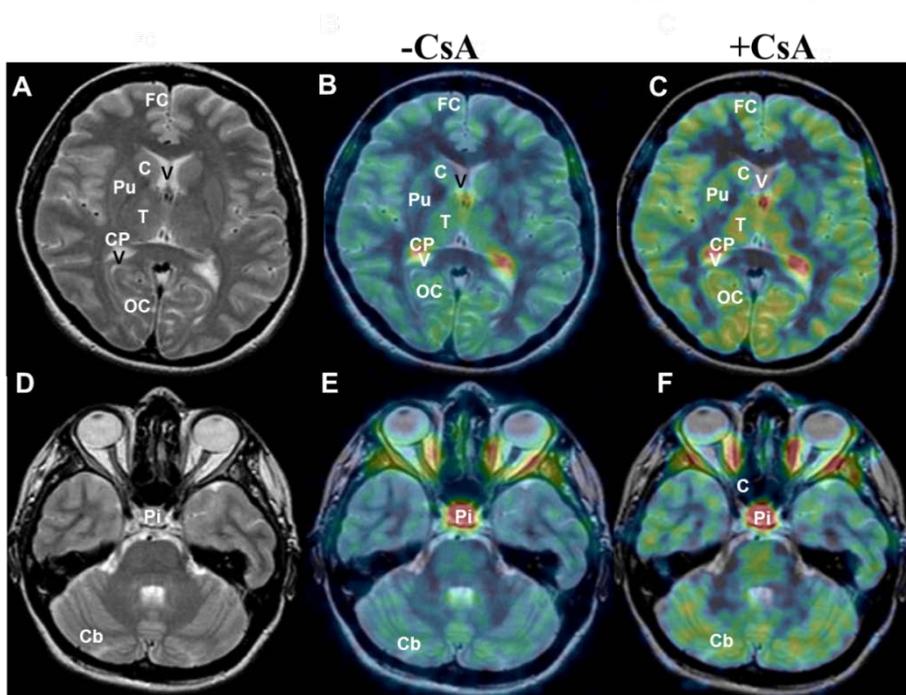
Eyal, Hsiao & Unadkat Pharmacol. Ther., 2009

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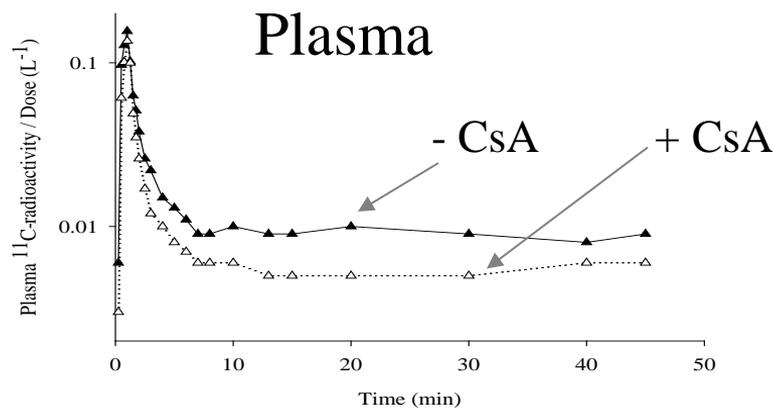
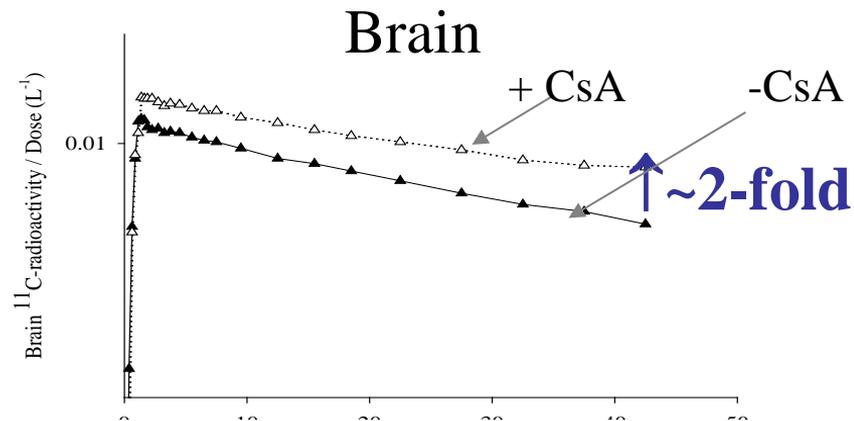
P-gp at the Rat BBB Results in Asymmetry in Brain:Blood Conc. of Verapamil (P-gp substrate)



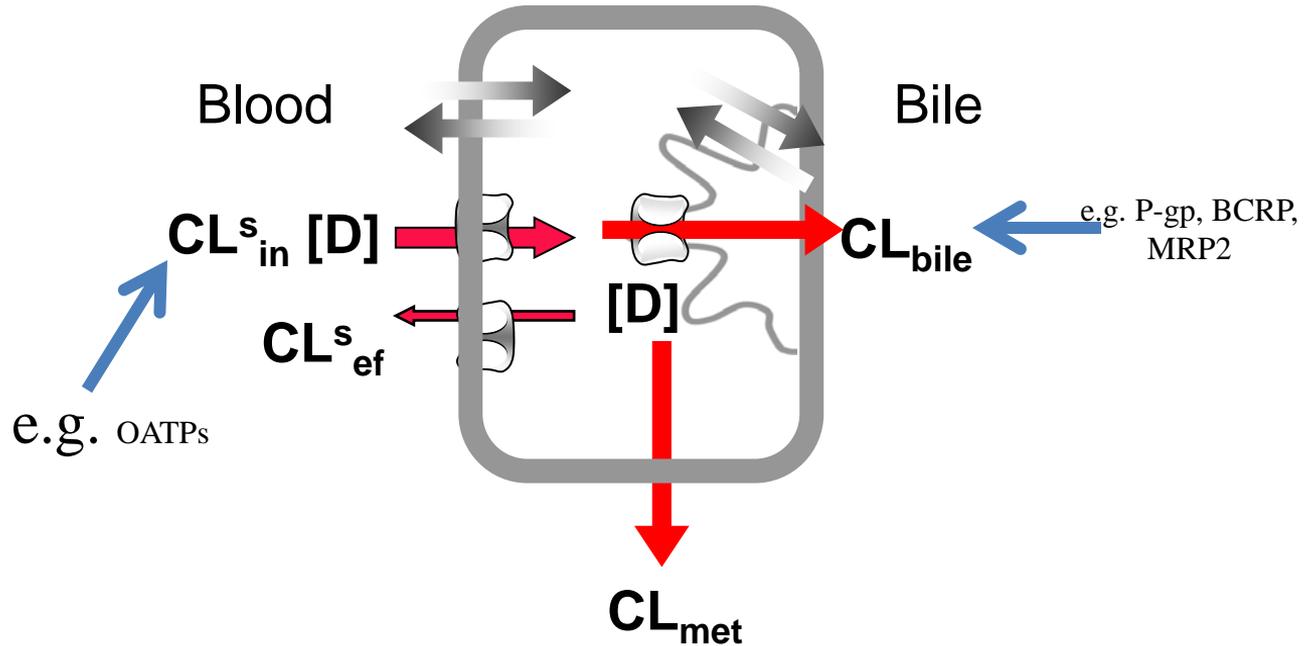
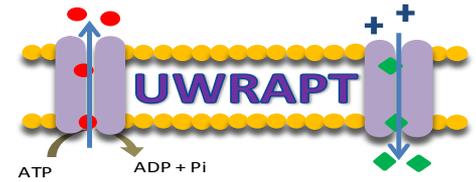
Asymmetry in Drug Conc. at the Human Brain:Blood Barrier: P-gp Efflux of ^{11}C -Verapamil



^{11}C -verapamil $\text{AUC}_{\text{brain}}:\text{AUC}_{\text{blood}}$ (20 min) - 0.42 ± 0.04



Asymmetry in Tissue: Blood Drug Conc. - Liver



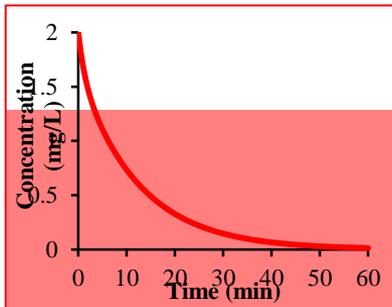
Asymmetry in Liver Tissue: Plasma Conc. when CL_{in}^S is the Rate-Determining Step

i.e. $CL_{met+bile} \gg CL_{ef}^S$

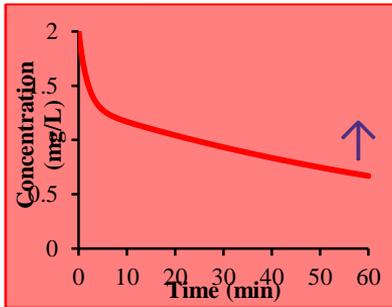


Blood

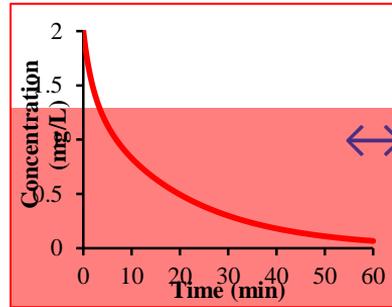
No inhibition



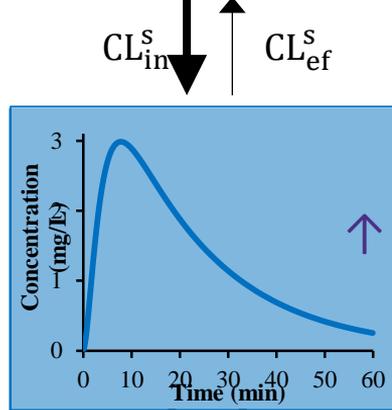
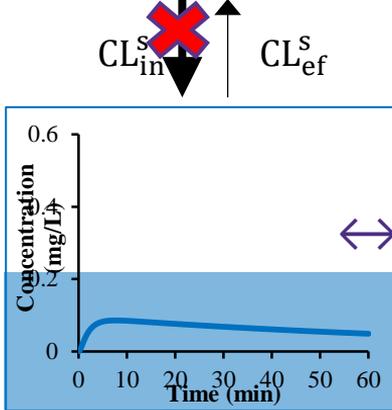
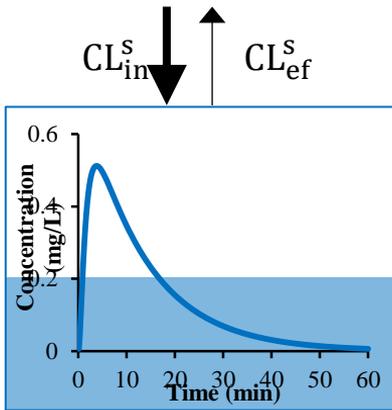
Inhibition of sinusoidal uptake CL



Inhibition of metabolism and biliary efflux CL



Liver



Assumptions:

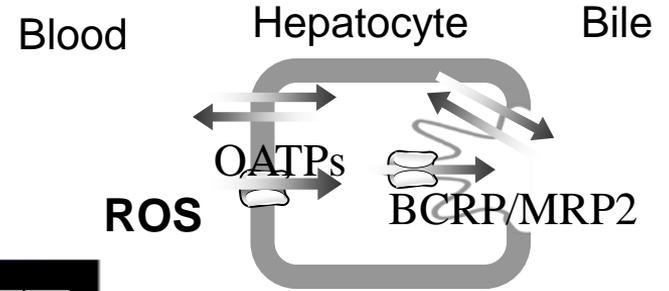
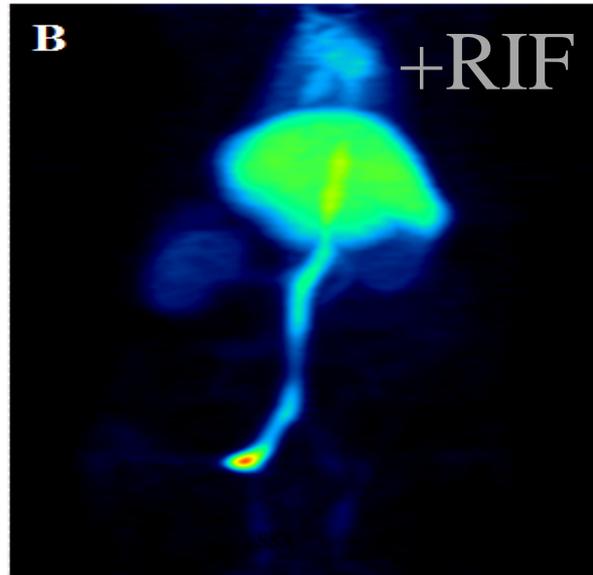
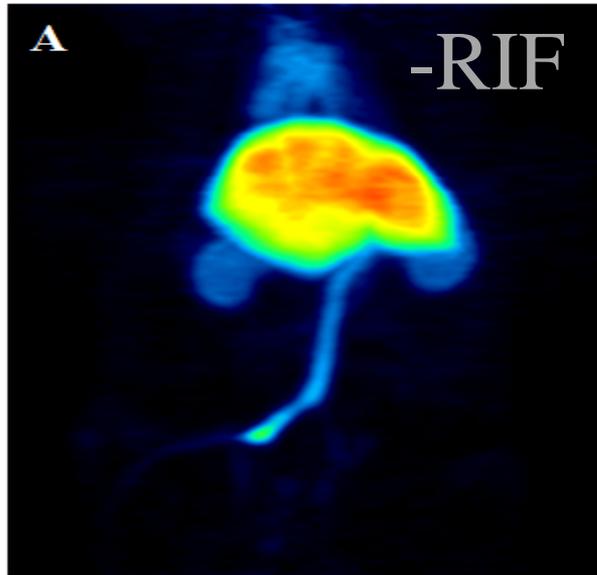
- > Liver is the only eliminating organ
- > 90% inhibition

Patilea-Vrana & Unadkat. CPT. 2016

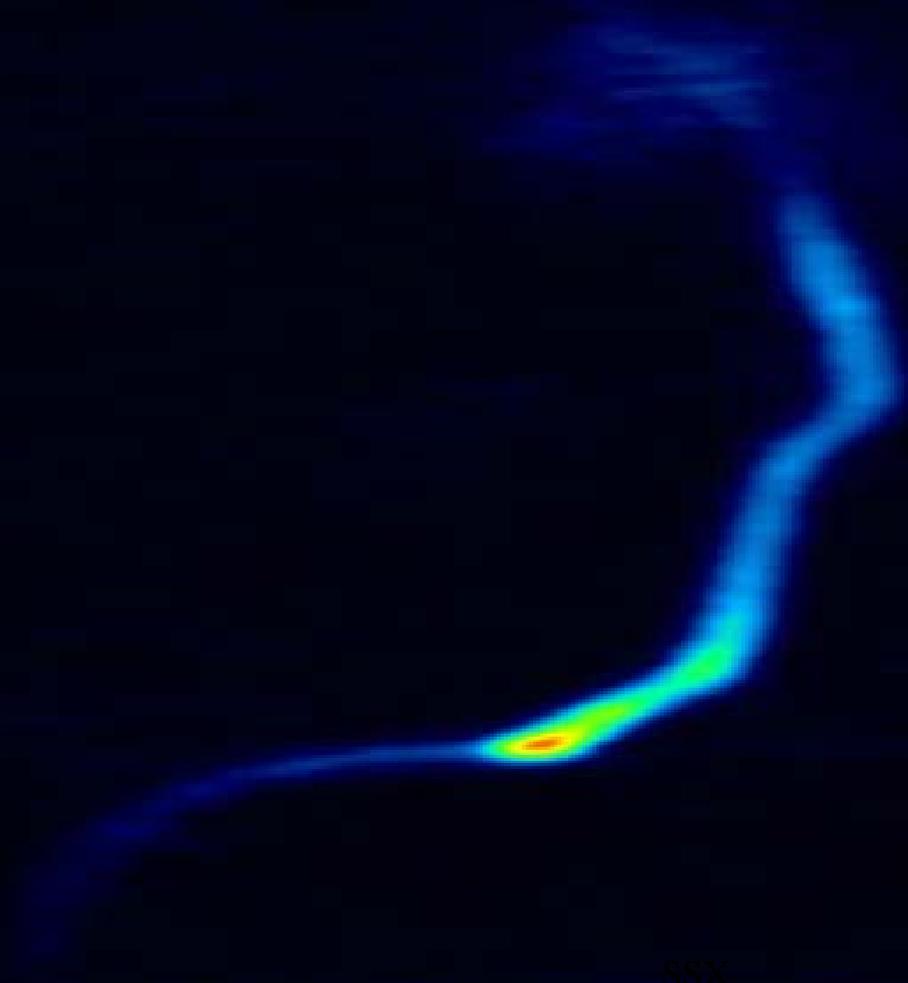
Asymmetry in Hepatic:Blood Conc. of ^{11}C -Rosuvastatin in the Rat



Coronal 2 min SUV images of ^{11}C -Rosuvastatin



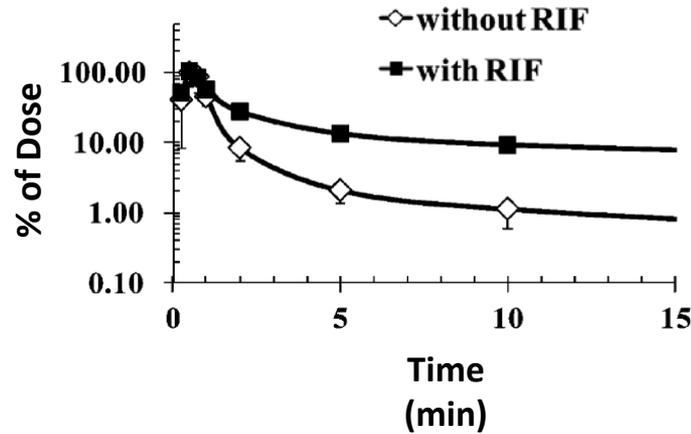
He et al., Mol Pharm., '14



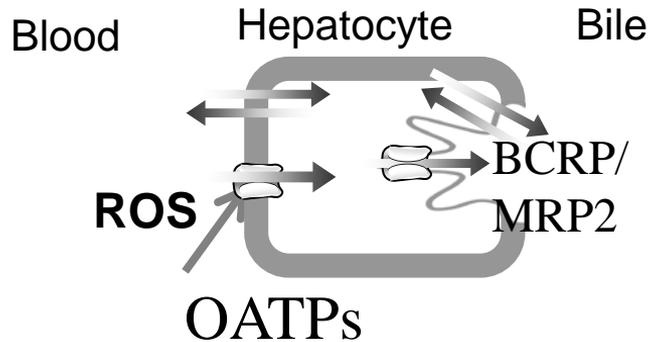
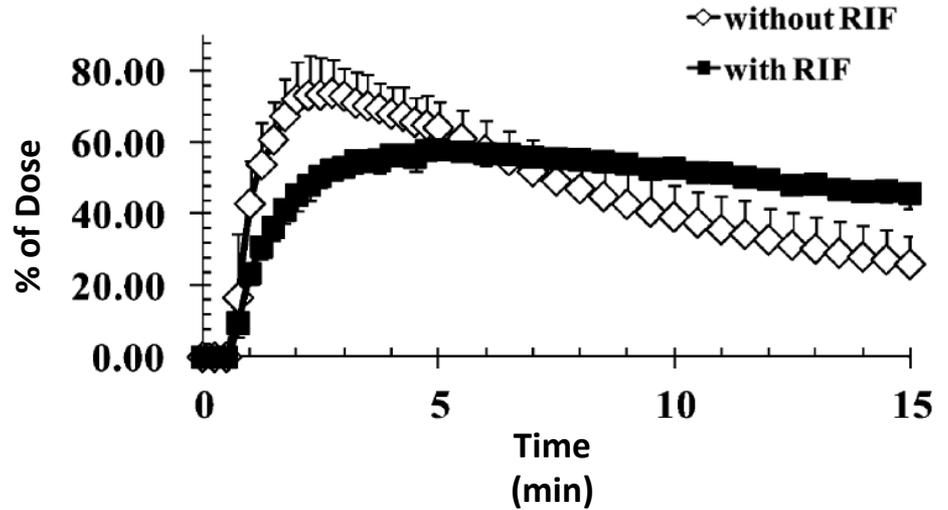
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Changes in Rat Liver Exposure to Rosuvastatin \pm Rifampin

Rat Blood

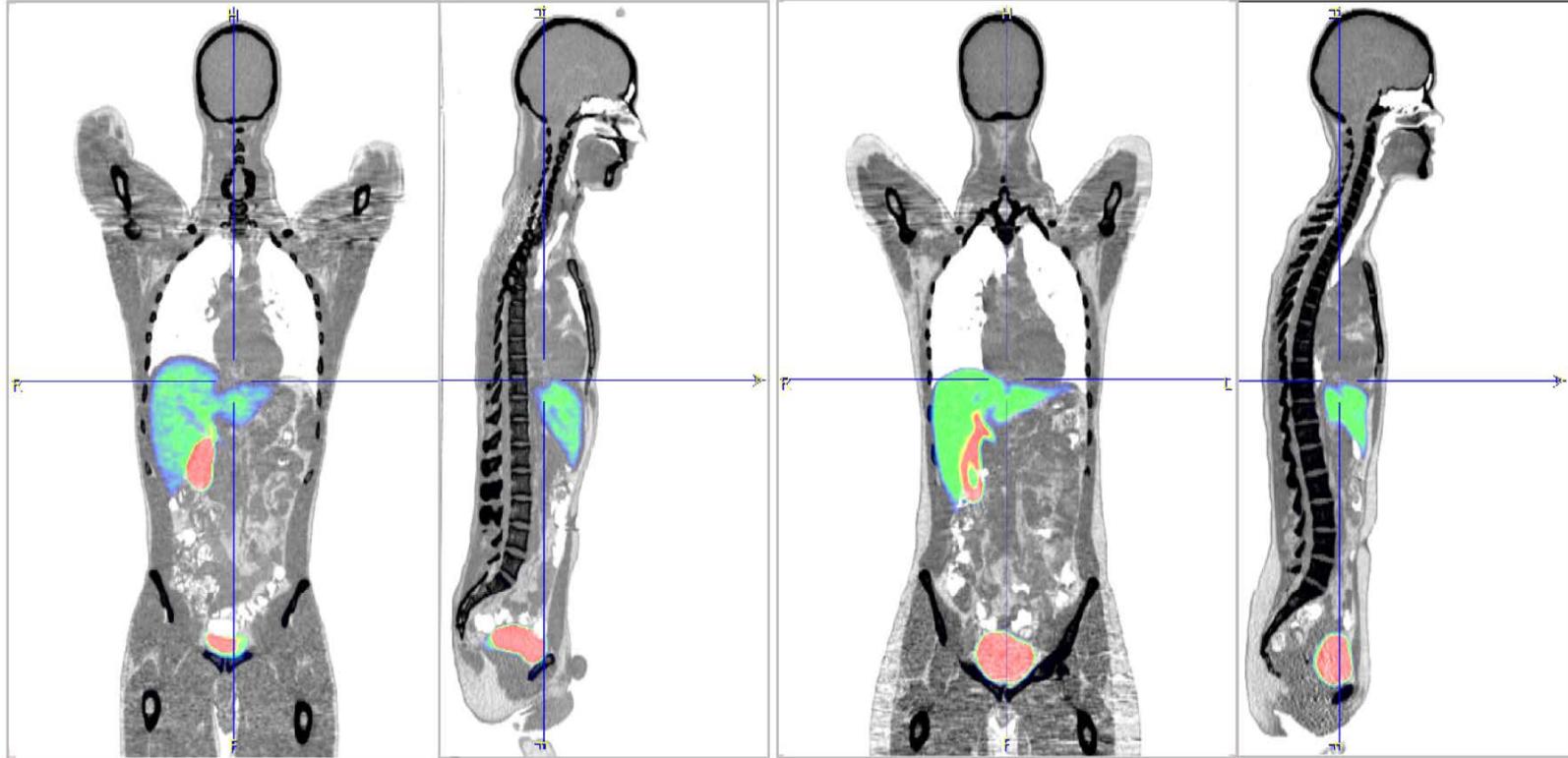
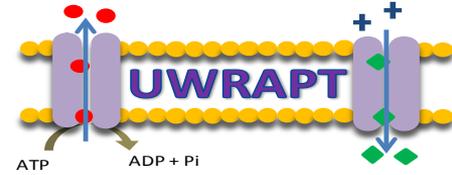


Rat Liver



2.3 fold increase in RSV plasma AUC but
NO significant increase in RSV liver AUC

Biodistribution of ^{11}C -rosuvastatin in humans



Billington et al., In progress

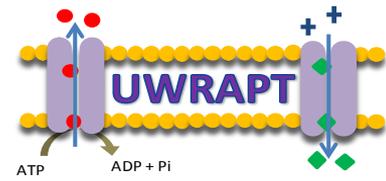
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How Can we Predict Tissue Drug Conc. in Humans?

- PET imaging (MRI and other imaging modalities do not have the required sensitivity):
 - Requires sophisticated equipment and radiochemistry
 - Costly (about \$20-40K/experiment/subject)
- Therefore we need alternative methods that will allow us to predict tissue conc. of drugs in humans

***APPLICATION OF PROTEOMICS DATA TO
PREDICT PK AND TISSUE CONC. OF DRUGS***

Hypothesis: Predict transporter-mediated *in-vivo* CL and tissue concentration of drugs in humans from *in-vitro* studies



Transporter-expressing cells



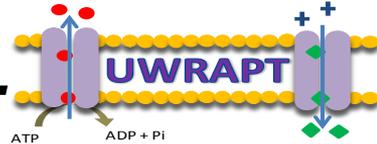
1. transporter-mediated drug CL
2. transporter abundance using quantitative proteomics
3. Obtain transporter scaling factor

Predict *in-vivo* CL and tissue conc. using transporter scaling factor

Verify predictions using PET imaging

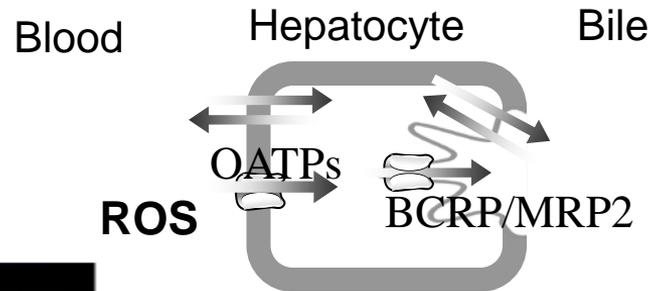
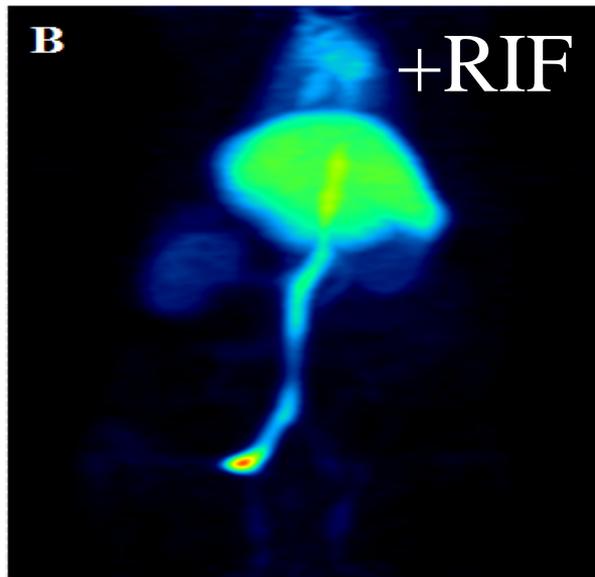
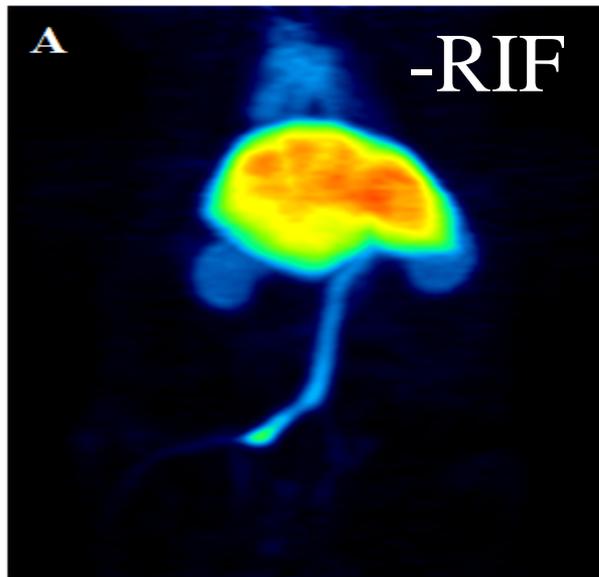


Hepatic Uptake and Biliary Excretion of ^{11}C -Rosuvastatin in the Rat



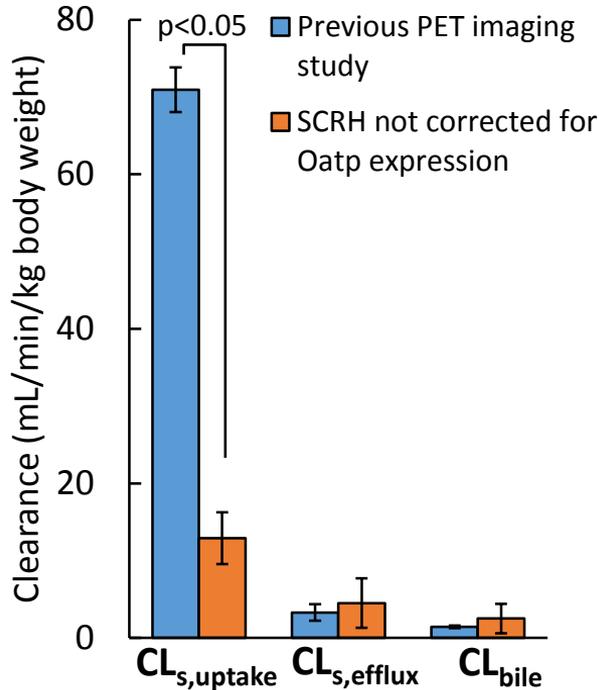
Coronal 2 min SUV images of ^{11}C -Rosuvastatin

0  10

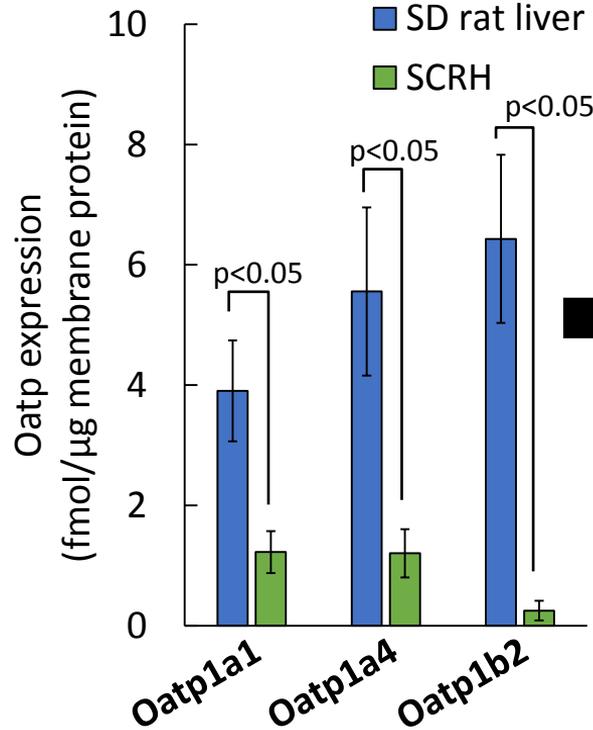


He et al., Mol Pharm., '14

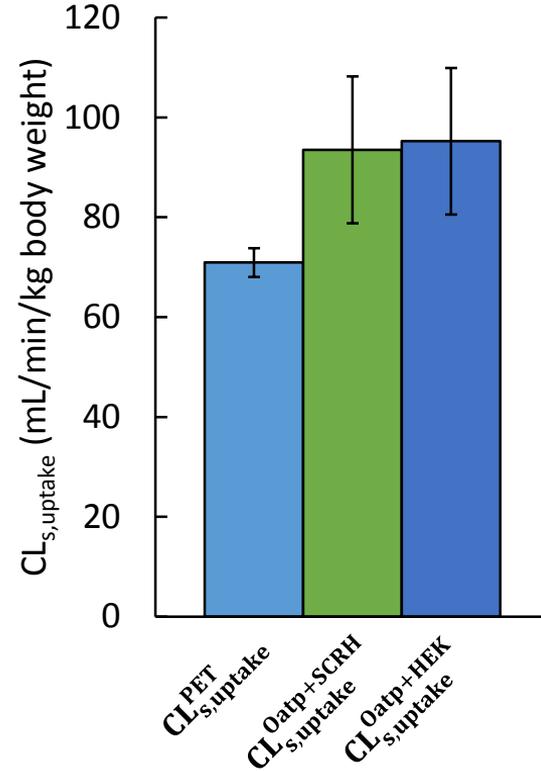
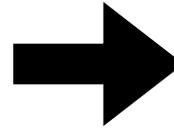
Successful prediction of the hepatobiliary clearance of rosuvastatin using cell lines, sandwich-cultured rat hepatocytes and quantitative proteomics



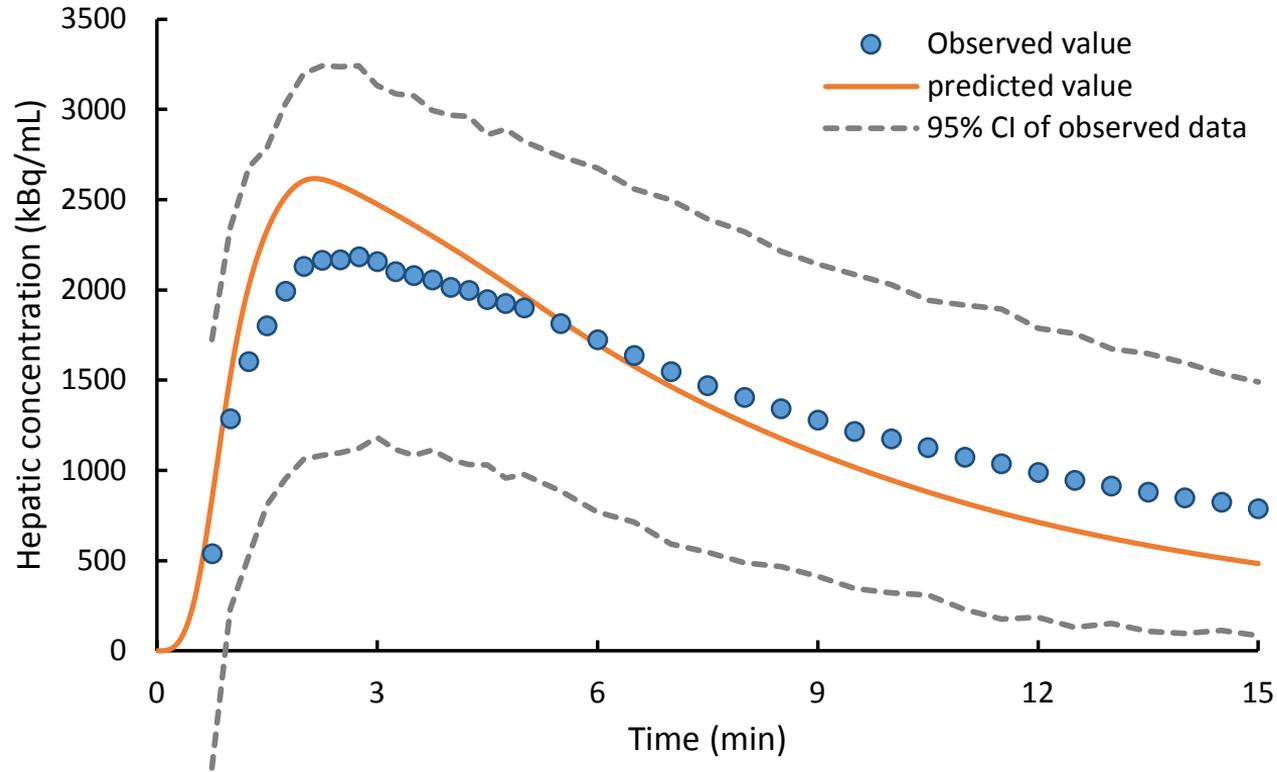
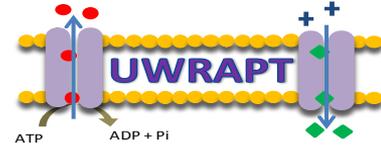
$CL_{s,uptake}$: sinusoidal uptake
 $CL_{s,efflux}$: sinusoidal efflux
 CL_{bile} : canalicular efflux



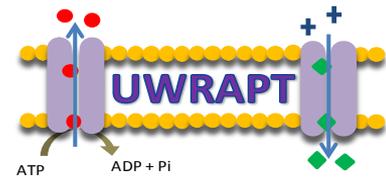
Ishida et al., DMD, 2018



Rat Hepatic Rosuvastatin Conc. well Predicted



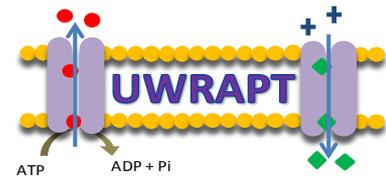
Can Rosuvastatin Hepatobiliary CL and Hepatic Conc. be Predicted in Humans?



Data removed as not published

Total transporter abundance in suspended (SH), plated (PH), sandwich-cultured (SCH) hepatocytes and liver tissue

Kumar et al.,
Unpublished data

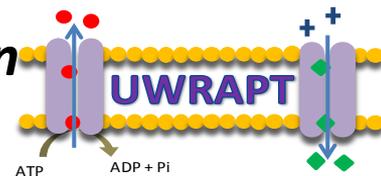


Data removed as not published

*Plasma membrane
transporter
abundance in
suspended (SH),
plated (PH),
sandwich-cultured
(SCH) hepatocytes
cf liver tissue*

Kumar et al.,
Unpublished data

Transporter-expressing cells better predict in-vivo (IV) human rosuvastatin hepatic uptake clearance than hepatocytes



Data removed as not published

$$CL_h = \frac{Q_h \cdot f_u \cdot CL_{s,uptake}}{Q_h + f_u \cdot CL_{s,uptake}}$$

*Assuming sinusoidal uptake is the rate determining step in RSV plasma CL

Patilea-Vrana G and Unadkat JD, *Clin Pharmacol Ther*, 2016

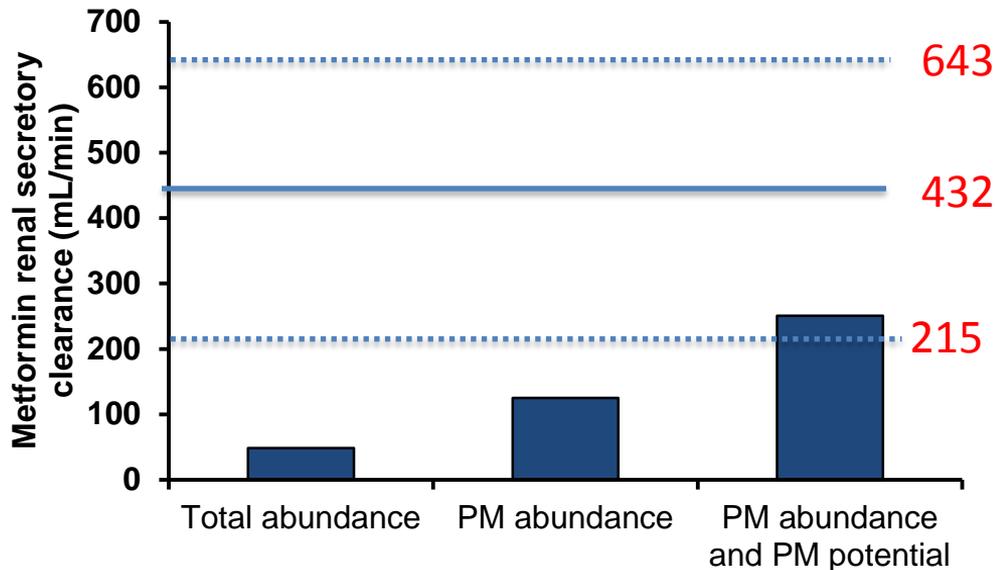
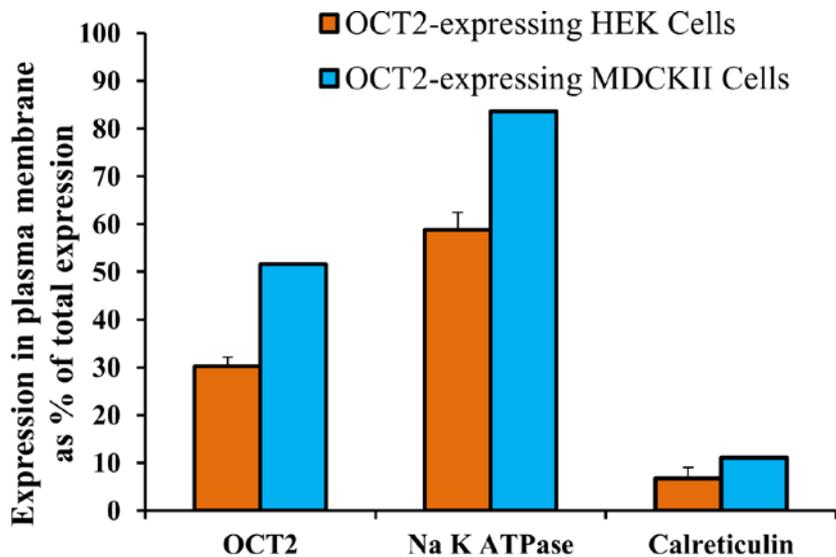
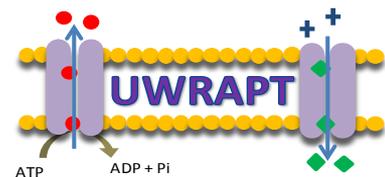
RSV uptake CL scaled on the basis of hepatocellularity *(Classical method)

*88 mg protein/g human liver

Martin et al., *Clin Ther*. 2003 Oct;25(10):2553-63.

Karlgren et al., *J Med Chem*. 2012 May 24;55(10):4740-63.

Metformin renal clearance is reasonably well-predicted using OCT2 expressing cells



Observed metformin renal secretory clearance in humans: 432 (range 215-643) mL/min

Summary



- Predicting tissue concentration and therefore efficacy and toxicity of a drug is the next frontier in ADME research
- The hepatic ECL model clarifies when transporters will or will not affect the systemic and tissue PK of a drug
- Tissue conc. measurement is possible using PET. However, this method cannot be routinely applied
- IVIVE using transfected cells and quantitative transporter proteomics is a promising technique to predict tissue drug conc.
- These predictions should be validated using PET imaging probes that interrogate multiple drug transporters

Major Contributors



Gabriela Patilea-Vrana



Sarah
Billington



YuYang

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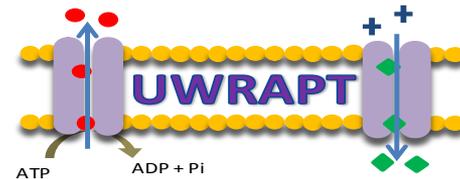
Vineet Kumar



Kazuya Ishida

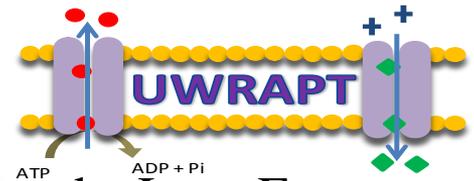


Bhagwat Prasad Anand Deo



Genentech, Merck,
Biogen, Gilead, BMS,
Takeda, Pfizer





Other Collaborators

Dept. of Radiology: Jeanne Link, David Mankoff, Todd Richards, Janet Eary, Satoshi Minoshima, Ken Maravilla, Mark Muzi, Steve Shoner, David Lewis, Jean Lee and the PET suite team

Dept. of Medicine: Ann Collier and her team; Scott Lee and his team

Dept. of Anesthesiology: Karen Domino, Matthew Pennington

Dept. of Pharmaceutics: Bhagwat Prasad, Edward Kelly, Carol Collins, Joanne Wang

Kidney Research Institute: Jonathan Himmelfarb

Univ. of Greifswald: Stefan Oswald and team

Children's Mercy Hospitals: Steven Leeder and team

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